Metaproteomics applied to activated sludge for industrial wastewater treatment revealed the dominance of Hyphomicrobium zavarzinii

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kDa

—250

—100

Introduction

In biological wastewater treatments microbial populations often cooperate in degradation of pollutants present in wastewater representing the biomass of the so-called activated sludge. In this work the metabolic behavior of the biomass of a lab-scale plant treating industrial pharmaceutical wastewater was investigated. Figure 1 shows the complete treatment process which included a membrane biological reactor (MBR) coupled with an advanced oxidation process (AOP) for partial breakdown of non-biodegradable molecules [1]. Proteins from biomass samples collected pre- and post-AOP application were investigated by two-dimensional gel electrophoresis (2DE) and MS then identified by database search. Results showed methanol dehydrogenase (MDH) belonging to Hyphomicrobium zavarzinii as the most constantly expressed protein in the studied consortium. Other constant identified proteins belonging to Hyphomicrobium spp. (figure 2) revealed a predominant methylotrophic metabolism, particularly H. zavarzinii appeared as key actor in the studied microbial community.

250 **—**

100 _

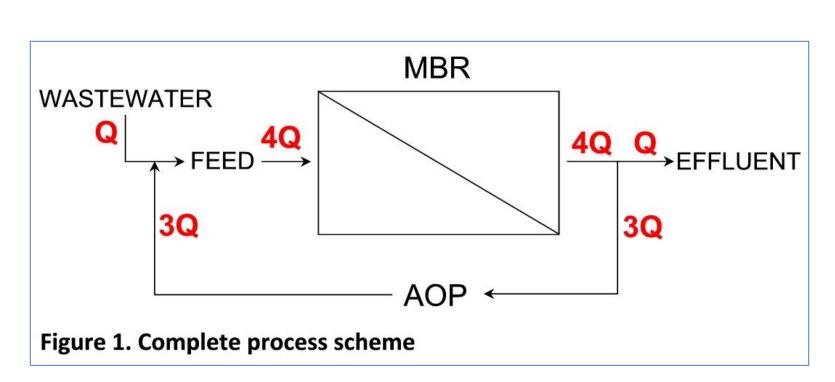
50 **–**

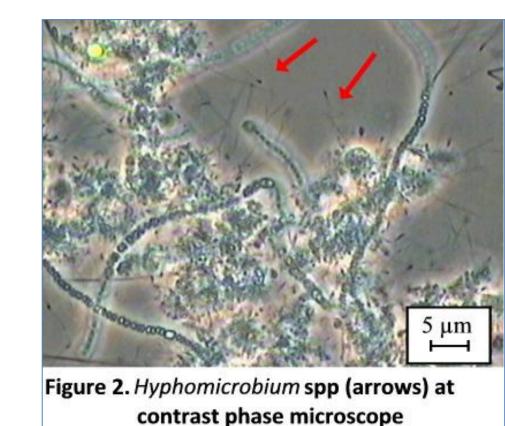
37_

25_

15 ___

10 __





Methods

Proteins were extracted as described elsewhere [2] and resolved by 2DE: first dimension was carried out through isoelectric focusing (IEF) by 18 cm IPG strips pH 4-7 (GE Healthcare), the second one through SDS-PAGE [3]. Delta2DTM software (Decodon, Greifswald, Germany) was used for image analysis. Spots of interest were manually excised, subjected to tryptic digestion and analyzed by MALDI TOF MS/MS (Ultraflextreme, Bruker Daltonics, Massachusetts, U.S.). Proteins were identified by searching peptide spectra with MASCOT (http://www.matrixscience.com) against NCBI.

Spot	gi of best hit	MW [kDa]	pl	MOWSE sc.	#Pept.	SC [%]	Functional group	Protein
1	518930489	69.8	6.41	295.25	4	8.2	Metabolism	quinoprotein ethanol dehydrogenase [Hyphomicrobium zavarzinii]
2	518930489	69.8	6.41	436.82	5	10.2	Metabolism	quinoprotein ethanol dehydrogenase [Hyphomicrobium zavarzinii]
3	518931602	69.0	6.07	566.66	7	14.6	Metabolism	methanol dehydrogenase [Hyphomicrobium zavarzinii]
4	518931602	69.0	6.07	405.02	8	15.1	Metabolism	methanol dehydrogenase [Hyphomicrobium zavarzinii]
5	338737333	69.5	5.80	629.00	9	18.1	Metabolism	methanol dehydrogenase subunit alpha [Hyphomicrobium sp. MC1]
6	518931602	69.0	6.07	605.55	10	17.2	Metabolism	methanol dehydrogenase [Hyphomicrobium zavarzinii]
7	518931602	69.0	6.07	313.37	7	10.5	Metabolism	methanol dehydrogenase [Hyphomicrobium zavarzinii]
8	563688372	17.9	5.50	295.54	2	17.6	Metabolism	aldehyde-activating protein [Hyphomicrobium nitrativorans NL23]
9	563688372	17.9	5.50	479.65	4	36.5	Metabolism	aldehyde-activating protein [Hyphomicrobium nitrativorans NL23]
10	563688372	17.9	5.50	556.55	5	45.3	Metabolism	aldehyde-activating protein [Hyphomicrobium nitrativorans NL23]

Table 1. Main protein identifications.

The main part of spots resulted constantly abundant from image analysis of 2DE. From overall 144 spots, 74 were analyzed by MS, and someone have been excluded for low Mowse score (less than 100) or insufficient number of peptides (less than 2 peptides). In conclusion 40 spots were reliably identified by MALDI TOF MS/MS: 36 constantly abundant spots and 4 spots with changing abundance. Methanol dehydrogenase (MDH) resulted the more expressed protein, ethanol dehydrogenase in methylotrophic metabolism. Metaproteome analysis showed that 31 database hits of identified proteins were denitrificans. Interestingly AAP presents the same domain of formaldehyde-activating enzyme (FAE) belonging to Methylobacterium extorquens AM1 (KEGG entry W911_13190), and Chistoserdova and coworkers showed the strong

Figure 3. 2D gels with main identified spots. Pre- and post-AOP gels are respectively on the left and on the right.



(EDH) and aldehyde-activating protein (AAP) were found constant too (figure 3 and table 1). All these proteins are involved taxonomically affiliated in the genus Hyphomicrobium: 17 particularly to H. zavarzinii, 9 to H. nitrativorans and 4 to H. relationship between MDH and FAE using methanol as source of energy and carbon [4].

References

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- 2.Kuhn R., Benndorf D., Rapp E., Reichl U., Palese L.L., Pollice A. (2011) Metaproteome analysis of sewage sludge from membrane bioreactors. Proteomics 11:2738-2744.
- 3.Laemmli U.K. (1970) Cleavage of structural proteins during the assembly of the head of Bacteriophage T4. Nature 227:680-685.
- 4. Chistoserdova L., Laukel M., Portais J.C., Vorholt J.A., Lidstrom M.E. (2004) Multiple formate dehydrogenase enzymes in the facultative methylotroph Methylobacterium extorquens AM1 are dispensable for growth on methanol. J. Bacteriol. 186:22-28.

Conclusions

The high and constant expressions of MDH and EDH proposed the methylotroph H. zavarzinii as principal player in the biomass. The identification of AAP of H. nitrativorans leads to the hypothesis of a metabolic cooperation with MDH of *H. zavarzinii* in the oxidation pathway of methanol. Further studies should be addressed to different samples of wastewater treatment, and could open new perspectives and scenarios towards novel possible applications of activated sludge biological consortia.



